DOI: http://dx.doi.org/10.18782/2320-7051.7320

**ISSN: 2320 – 7051** *Int. J. Pure App. Biosci.* **7 (2):** 98-104 (2019)

**Research** Article



## Enzymatic Treatment (a galactosidase) to Reduce Antinutrient Content in Raw Jackfruit Bulb Flour and Seed Flour

Anila, H. L. and Suma Divakar<sup>\*</sup>

Department of Community Science, College of Agriculture, Vellayani, Thiruvananthapuram, Kerala, India \*Corresponding Author E-mail: divakarsuma67@gmail.com Received: 26.02.2019 | Revised: 30.03.2019 | Accepted: 6.04.2019

### ABSTRACT

Oligosaccharides are reported to be the cause of flatulence in jackfruit. With the aim of reducing these antinutritional factors, ' $\alpha$  galactosidase' enzyme was utilised to breakdown oligosaccharides to simpler compounds and hence reduce the indigestible factors. Koozha type jackfruit in their raw stage was selected. Bulbs and seeds were separated and processed into flour. Flours were made from dough to batter by adding water and subjected to enzymatic treatment. The enzyme ' $\alpha$  galactosidase' was premixed with the flour of jackfruit seed and jackfruit bulb separately in the ratio 1:100 and the moisture level was varied from 25 – 200% (dough to batter stage). The hydrolysis was carried out for 90 minutes in both jackfruit bulb flour and seed flour. The products were evaluated for the breakdown of oligosaccharides (Raffinose) using HPTLC method. The results showed that the level of Raffinose after treatment with bulb flour and seed flour was seen to decrease with increase in moisture content (25-100%). Thereafter the content slightly staggered and then reduced (125%, 150%, 175%, 200%). Overall the level of oligosaccharide decreased in comparison to levels in control (0.97  $\mu$ g g<sup>-1</sup>).

*Key words:* Enzymatic treatment,  $\alpha$  galactosidase, Oligosaccharides, Raffinose, Jackfruit bulb flour, Jackfruit seed flour, Moisture level

#### **INTRODUCTION**

Anti-nutrients may occur naturally in plants as secondary metabolites, protecting plants from viral and fungal attack analogous to the immune system of animals. Anti-nutrients may also be produced in large amounts, as a direct result of some adverse environmental condition<sup>7</sup>.

Robin and Ross<sup>17</sup> highlight that antinutrients are not something to be alarmed, since most foods typically have one or more antinutrients and that the issue is their concentration and type and whether that specific antinutrient profile will adversely affect health. All plants have some antinutrient properties, soybean plant is especially rich in these chemicals. If they are not removed by extensive preparation such as fermentation or soaking, soybeans can become one of the worst foods a person can eat<sup>16</sup>.

Oligosaccharides of the raffinoseseries (namely raffinose, verbascose and stachyose) are major components in many foods.

**Cite this article:** Anila, H.L. and Divakar, S., Enzymatic Treatment (α galactosidase) to Reduce Antinutrient Content in Raw Jackfruit Bulb Flour and Seed Flour, *Int. J. Pure App. Biosci.* **7**(2): 98-104 (2019). doi: http://dx.doi.org/10.18782/2320-7051.7320

ISSN: 2320 - 7051

Raffinose-series of oligosaccharides are not hydrolysed in the upper gut due to the absence of  $\alpha$ -galactosidase. In the lower intestine they are metabolised by bacterial action, producing methane, hydrogen and carbon dioxide, which lead to flatulence and diarrhoea. Raffinoseseries oligosaccharides are thus a major factor limiting the use of grain legumes in monogastric diets. Flatulence is a complaint even among healthy individuals and is one of the common causes of abdominal discomfort. It is also associated with dyspepsia, constipation and diarrhoea<sup>14</sup>.

Raffinose family oligosaccharides (RFOs) are complex sugars containing chains of ' $\alpha$  galactose' which are unable to be digested in the human upper intestine, due to the absence of  $\alpha$  galactosidase, the enzyme required to break the links in the  $\alpha$  galactose chains. The RFOs therefore survive through to the lower intestine, where they encounter bacteria which use them as the substrate for fermentation. This produces methane, carbon dioxide and hydrogen gas, which can cause bloating, abdominal discomfort and excessive flatulence<sup>2,3,10,18</sup>.

### MATERIAL AND METHODS

### Treatment with enzyme α galactosidase

The enzyme  $\alpha$ -Galactosidase catalyzes the hydrolysis of  $\alpha$ -1,6-linked galactoside residues

in simple oligosaccharides, such as melibiose, raffinose, and stachyose and the polysaccharides of galactomannans and guar gum. It also acts on glycoconjugates, glycoproteins and glycosphingolipids and is known catalyze transglycosylation to reactions, especially with higher substrate concentrations<sup>19</sup>. Thus, interest in this enzyme stems from its potential biotechnological and medicinal applications. Currently, the most important industrial applications of αgalactosidase are related to the beet sugar industry, paper pulp industry, soya food processing and animal feed processing<sup>15</sup>.

The enzyme  $\alpha$  galactosidase was premixed with the dry flour of jackfruit seed and jackfruit bulb separately in the ratio 1:100<sup>4</sup>. The moisture level of the flour was varied from 25 – 200% (dough to batter stage). The hydrolysis was carried out for 90 minutes in both jackfruit bulb flour and seed flour. The products were evaluated for the breakdown of oligosaccharides. The details of variations in moisture content are explained below.

### Statistical Design and treatment details

Experimental design : Completely randomized design (CRD) Number of treatments : 8 x 2 Number of replications : 2

Treatments	Moisture level (%)
$\mathbf{M}_1$	25
$M_2$	50
$M_3$	75
$M_4$	100
$M_5$	125
$M_6$	150
$M_7$	175
$M_8$	200

### **Edible Portion**

E<sub>1</sub> Bulb (flour)

 $E_2$  Seed (flour)

# HPTLC estimation of Oligosaccharides (Raffinose $\mu g g^{-1}$ )

HighPerformanceThinLayerChromatography(HPTLC)isacommonly

used method for the analysis of food bioactive oligosaccharides because it is available in most of laboratories and does not require much technical support. This method is popular due to its simplicity and adaptability and the availability of the equipment.

ISSN: 2320 - 7051

### Anila and Divakar

The concentration of Oligosaccharides (Raffinose) were analysed for enzyme treated flours of jackfruit bulbs and seeds, through High Performance Thin Layer Chromatography (HPTLC) analysis.

Raffinose (reference standard of 99 per cent purity) was procured from Sigma – Aldrich Chemie GmbH (Aldrich Division; Steinheim, Federal Republic of Germany). Isopropanol (purity 99 per cent), methanol (purity 99 per cent), toluene(purity 99 per cent), and ammonia solution (95 per cent) were procured from Merck, India. All solvents were of HPLC grade and distilled water used was purified with Sartorius water purification unit (Arium 61315, made in USA).

### **Extraction of samples for HPTLC**

Methanolic extract of the treated and pre treated jackfruit bulb flour and seed flour was obtained by collecting filtrate (Whatmann No.41 filter paper) of ten gram powder added with 25 ml methanol, shaken in a rotary evaporator (30 rpm) over night at room temperature.

### **Mobile Phase**

Mobile phase selected for detection of oligosaccharide (Raffinose) was a mixture of isopropanol, methanol and toluene in the ratio of 8: 2: 4 and ammonia (1 drop), sonicated for 10 minutes.

# Stock Solution of Oligosaccharide (Raffinose)

Stock solution of Raffinose  $(1000 \ \mu g \ ml^{-1})$  was prepared by mixing 10 mg Raffinose reference standard and five millilitre methanol by thoroughly shaking it in a volumetric flask (10 ml) and making up to the mark using methanol.

# Working Standard Solution of Oligosaccharide (Raffinose)

Working standard solution (1 to 30  $\mu$ g ml<sup>-1</sup>) was prepared by obtaining the aliquots (0.01 – 0.30 ml) from the stock solution of Raffinose and each of its volume made up to 10 ml with methanol.

### **Operating System Conditions**

In HPTLC analysis, the samples were spotted in the form of bands of width six millimetre, which were 14 millimeter apart with a Camag microlitre syringe on pre- coated silica gel aluminium plate (60 F254 having 20x 10 cm). Linear ascending development was carried out in 20x10 cm twin trough glass chamber (Camag, Muttenz, Switzerland) saturated with the mobile phase. The optimized chamber saturation time for mobile phase was 20 minutes at room temperature. The length of chromatogram run was 80 mm. Densitometric scanning was performed on Camag TLC scanner III in the reflectance fluorescence mode at 254 nm, as well as 366 nm and was operated by CATS software (V3. Camag).

### **RESULTS AND DISCUSSION** Reduction of oligosaccharide (Raffinose) content in bulb flour

The oligosaccharide (Raffinose) content in pre treated and treated jackfruit bulb flour (JFBF) at different moisture levels differed customarily in the HPTLC assays. In HPTLC analysis, the retention factor of standared Raffinose was 0.58 minutes. The distance that each component of a mixture travels can be quantified using retention factors (Rf). The retention factor of a particular material is the ratio of the distance the spot moved above the origin to the distance the solvent front moved above the origin. Retention factor recorded by jackfruit bulb flour (control) and bulb flour with enzyme treated at varying moisture levels were 0.58, 0.58, 0.58, 0.58, 0.58, 0.57, 0.57, 0.57, 0.57 at 25%, 50%, 75%, 100%, 125%, 150%, 175%, 200% respectively (Table 1).

These values were compared to that of the standard Raffinose. Raffinose content in jackfruit bulb flour was 0.97in control (without treatment); while the amount in treated samples were in 0.89 in M<sub>1</sub>, 0.85 in M<sub>2</sub>, 0.80 in M<sub>3</sub>, 0.71 in M<sub>4</sub>, 0.74 in M<sub>5</sub>, 0.83 in M<sub>6</sub>, 0.75 in M<sub>7</sub>, 0.82 $\mu$ g g<sup>-1</sup> in M<sub>8</sub> respectively. The level of Raffinose after treatment was seen to decrease with increase in moisture content (25-100%). Thereafter the variation was not uniform (as 0.74 in M<sub>5</sub>, 0.83 in M<sub>6</sub>, 0.75 in M<sub>7</sub> and 0.82 in M<sub>8</sub>) but the level of oligosaccharides decreased in comparison to control (0.97  $\mu$ g g<sup>-1</sup>), which indicates that the enzyme was effective in breakdown of

oligosaccharides. The enzyme  $\alpha$  galactosidase is reported to have the ability to hydrolyse stachyose and raffinose<sup>1,5,9,8,12</sup>.

# Reduction of oligosaccharide (Raffinose) content in seed flour

The oligosaccharide (Raffinose) content in pre treated and enzyme treated jackfruit seed flour (JFSF) at different moisture levels differed in HPTLC assay and the data is tabulated in Table 2. In the HPTLC analysis, the retention factor of standard Raffinose was 0.61 minutes. Retention factor recorded by pretreated (0.61 minutes) and treated jackfruit seed flour was compared to that of the standard Raffinose. Raffinose content in plain jackfruit bulb flour was 1.16 and 0.83, 0.68, 0.63, 0.56, 0.50, 0.58, 0.74 and 1.08  $\mu$ g g<sup>-1</sup> at 25%, 50%, 75%, 100%, 125%, 150%, 175%, 200% dilutions respectively. The level of Raffinose content in seed flour after treatment was seen to decrease with increase in moisture content (25-100%). Thereafter the variation was not uniform (125%, 150%, 175%, 200%). However it may be noted that the level of oligosaccharides decreased in comparison to control (1.16  $\mu$ g g<sup>-1</sup>), which indicates the asymptotic effect of enzyme on breakdown of oligosaccharides.

Guimaraes et al.11, found that incubation of soymilk with  $\alpha$  galactosidase for 8 hours at 30°C resulted in a 73.3 per cent reduction in raffinose and a 40.6 per cent reduction in stachyose. In another study conducted by Matella et al.<sup>13</sup>, it was found that incubation of black, red and navy beans with  $\alpha$  galactosidase for 1 hour at 23°C was more effective (30%-50% reduction) in the reduction of RFOs than soaking of beans for 5 hours at 23°C (1%-35% reduction). Song and Chang<sup>20</sup> found that two hours of incubation with a galactosidase at 55-60<sup>°</sup>C removed 100 per cent of RFOs in pinto beans. This is a significantly greater reduction, than was achieved by soaking followed by boiling for 90 minutes (54.2%).

 Table 1: Quantification of Oligosaccharide (Raffinose) in jackfruit bulb flour at different moisture levels and enzyme treatment

Treatments	<b>Retention Factor</b>	Area	Amount (µg/g <sup>-1</sup> )
(%)		(AU)	
Standard	0.58	6319.7	-
M <sub>0</sub> (control)	0.58	6178.0	0.97
M <sub>1</sub> 25	0.58	5677.3	0.89
M <sub>2</sub> 50	0.58	5394.8	0.85
M <sub>3</sub> 75	0.58	5117.0	0.80
M <sub>4</sub> 100	0.58	4500.9	0.71
M <sub>5</sub> 125	0.57	4734.9	0.74
M <sub>6</sub> 150	0.57	5307.7	0.83
M <sub>7</sub> 175	0.57	4795.3	0.75
M <sub>8</sub> 200	0.57	5185.9	0.82

Table 2: Quantification of Oligosaccharide (Raffinose) in jackfruit seed flour at different moisture levels	
and enzyme treatment	

Treatments	Retention	Area	Amount
(%)	Factor	(AU)	(µg g <sup>-1</sup> )
Standard	0.61	5102.6	-
M <sub>0</sub> (control)	0.61	5963.0	1.16
M <sub>1</sub> 25	0.61	4265.2	0.83
M <sub>2</sub> 50	0.61	3510.5	0.68
M <sub>3</sub> 75	0.61	3253.0	0.63
M <sub>4</sub> 100	0.61	2866.2	0.56
M <sub>5</sub> 125	0.61	2595.2	0.50
M <sub>6</sub> 150	0.61	2986.6	0.58
M <sub>7</sub> 175	0.61	3824.3	0.74
M <sub>8</sub> 200	0.61	5557.7	1.08

III	Juis		
Treatments	Raffinose (µg g <sup>-1</sup> )		
Moisture level			
M <sub>0</sub>	1.07		
$M_1$	0.86		
<b>M</b> <sub>2</sub>	0.76		
M <sub>3</sub>	0.71		
$M_4$	0.63		
M5	0.62		
M <sub>6</sub>	0.70		
M <sub>7</sub>	0.74		
M <sub>8</sub>	0.95		
SEm <u>+</u>	0.001		
CD	0.004		
Edible portion			
E <sub>1</sub>	0.82		
E <sub>2</sub>	0.75		
SEm <u>+</u>	0.001		
CD	0.002		

 Table 3: One way interaction effect of moisture levels and edible portion on raffinose content of jackfruit flours

Table 3. shows the effect of varying moisture levels and edible portion on raffinose content (2 way interaction). From this table it is observed that the raffinose content was higher in control samples of jackfruit bulb flour and seed flour (1.07  $\mu$ g/g<sup>-1</sup>). The lowest raffinose content was observed in treatment M<sub>5</sub> (125% moisture level). With respect to edible portion, E<sub>1</sub> (Bulb flour) contained 0.82  $\mu$ g g<sup>-1</sup>and E<sub>2</sub>

(Seed flour) contained 0.75  $\mu$ g g<sup>-1</sup>. Dey and Pridham<sup>6</sup> observed that generally  $\alpha$ galactosidase may hydrolyze a variety of simple  $\alpha$ -d-galactosides as well as more complex molecules, such as oligosaccharides and polysaccharides.  $\alpha$ -Galactosidase ( $\alpha$ galactoside galactohydrolase EC 3.2.1.22) is widely distributed in microorganisms, plants and animals.

Table 4: Two way interaction effect of moisture level and edible portion on raffinose conte	nt

Treatments	Raffinose (µg g <sup>-1</sup> )
$M_0E_1$	0.97
$M_0E_2$	1.16
$M_1E_1$	0.89
$M_1E_2$	0.83
$M_2E_1$	0.85
$M_2E_2$	0.68
$M_3E_1$	0.80
$M_3E_2$	0.63
$M_4E_1$	0.71
$M_4E_2$	0.56
$M_5E_1$	0.74
$M_5E_2$	0.50
$M_6E_1$	0.83
$M_6E_2$	0.58
$M_7E_1$	0.75
$M_7E_2$	0.74
$M_8E_1$	0.82
$M_8E_2$	1.08
SEm <u>+</u>	0.002
CD	0.005

Int. J. Pure App. Biosci. 7 (2): 98-104 (2019)

#### ISSN: 2320 - 7051

Table 4 shows the two way interaction effect of varying moisture level and edible portion on raffinose content. Here it is observed that  $M_0E_2$  (Control seed flour) contained highest level of raffinose content (1.16 µg g<sup>-1</sup>), the lowest raffinose content was observed in  $M_5E_2$ (125% moisture level of seed flour), which contained 0.50 µg g<sup>-1</sup>. Brain<sup>4</sup> reported the dependence of enzyme  $\alpha$  galactosidase activity on the moisture content of the system with a steady increase in the proportion of both stachyose and raffinose hydrolysed with increasing moisture content. The results also showed a steady decrease in the concentration of sucrose and a steady increase in the concentration of galactose as moisture content increased. Sucrose and galactose are the hydrolysis products of stachyose and raffinose.

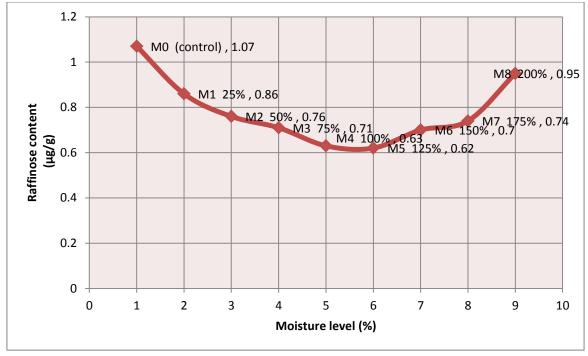


Fig. 1: Raffinose content with varying moisture levels

### CONCLUSION

The level of Raffinose content in seed flour after treatment was seen to decrease with increase in moisture content (25-100%). Here the raffinose content was seen to reduce up to 125%. Thereafter the variation was not uniform. The level of oligosaccharide however decreased in comparison to control (1.16  $\mu$ g g<sup>-1</sup>), which indicates the asymptotic effect of enzyme on breakdown of oligosaccharides.

### Acknowledgements

The authors place on record their whole hearted gratitude to Kerala Agricultural University, for the technical and financial support rendered and Ayurveda college, Trivandrum for support with the HPTLC facility.

#### REFERENCES

- Adya, S. and Elbein, A. D., Glycoprotein enzymes secreted by aspergillus niger: purification and properties of alpha galactosidase. *J. Bacteriol.* 850-856 (1977).
- Akinyele, I.O. and Akinlosotu, A., Effect of soaking, dehulling and fermentation on the oligosaccharides and nutrient content of cowpeas (*Vigna unguiculata*). *Food Chem.* 41: 43–53 (1991).
- Barampama, Z. and Simard, R. E., Effects of soaking, cooking and fermentation on composition, in-vitro starch digestibility and nutritive value of common beans. *Plants Foods Human Nutr.* 48: 349–365 (1994).

- Brain, C. J., Strategies for the removal of raffinose family oligosaccharides from navy bean flour. Ph.D Thesis, Massey University, Palmerston North, New Zealand. 110p. (2013).
- Civas, A., Eberhard, R., Dizet, P. L., and Petek, F., Glycosidases induced in *Aspergillus tamarii*: secreated at D galactosidases and f/D-mannanase. *Biochem. J.* pp.857-863 (1984).
- Dey, P.M. and Pridham, J.B., Biochemistry of α-galactosidase. Adv Enzymol. 36: 91-130 (1972).
- Duenas, M., Hernandez, T., Estrella, I., and Fernandez, D., Germination as a process to increase the polyphenol content and antioxidant activity of lupin seeds (*Lupinus angustifolius* L.). *Food Chem.* (117): 599-607 (2009).
- Ferreira, J.G., Reis, A. P., Guimaraes, V.M., Falkoski, D. L., Fialho, L. D. S., and Rezende, S. T. D., Purification and characterisation of Aspergillus terreus alpha galactosidases and their use for hydrolysis of soymilk oligosaccharides. *Appl. Biochem. Biotechnol.* 164: 1111-1125 (2011).
- Gherardini, F., Babcock, M., and Salyers, A. A., Purification and characterisation of two alpha galactosidase associated with catabolism of guar gum and other oa galactosides by bacteroides ovatus. *J. Bacteriol*.pp.500-506 (1985).
- Guillon, F. and Champ, M.M.J., Carbohydrate fractions of legumes: uses in human nutrition and potential for health. *Br. J. Nutr.* 88: 293-306 (2002).
- Guimaraes, V.M., Rezebde, S. T. D., Moreira, S. T., Barros, E. G. D., and Felix, C. R., Characterisation of alpha galctosidase from germinating soyabean seed and their use for hydrolysis of oligosaccharides. *Phytochem*.67-73 (2001).
- 12. Katrolia, P., Jia, H., Yan, Q., Song, S., Jiang, Z., and Haibo, X., Characterization of a protease resistant alpha galactosidase

from the thermophilic fungus *Rhizomucor miehei* and its application in removal of raffinose family oligosaccharides. *Bioresource Technol.* pp.578-586 (2012).

- Matella, N.J., Dolan, K.D., Stoeckle, A.W., Bennink, M.R., Lee, Y. S., and Uebersax, M. A., Use of hydration, germination and alpha galactosidase treatments to reduce oligosaccharides in dry beans. *J. Food Sci.* **70**: 203-207 (2005).
- Mitsou, E. K., Panopoulou, N., Turunen, K., Spiliotis, V., and Kyriacou, A., Impact of a jelly containing short-chain fructooligosaccharides and Sideritis euboea extract on human faecal microbiota. *Int. J. Food Microbiol.* 135: 112–117 (2010).
- Prashanth, S. J. and Mulimani, V. H., Soymilk oligosaccharide hydrolysis by Aspergillus oryzae [alpha]-galactosidase immobilized in calcium alginate. *Process Biochem.* 40: 1199-1205 (2005).
- Preet, K. and Punia, D., "Antinutrients and digestibility (in vitro) of soaked, dehulled and germinated cowpeas", *Nutrition and Health.* 14(2): 109-117 (2000).
- 17. Robin, D. G. and Ross, M. W., Breeding for staple food crops with high micronutrient density. pp.6-9 (1996).
- Shimelis, E.A. and Rakshit, S. K., Effect of processing on anti-nutrients and in vitro protein digestibility of kidney bean (*Phaseolus vulgaris* L.) varieties grown in East Africa. *Food Chem.* 103: 161-172 (2007).
- Silva, G. P., Moreno, A., Marques, F., Oliver, C., Jamur, M. C., Panunto, C. A., and Roque, M. C., Neutrophil activation induced by the lectin KM+ involves binding to CXCR2. *Biochem.* 1760(1): 86-94 (2006).
- Song, D. and Chang, S. K. C., "Enzymatic degradation of oligosaccharides in pinto bean flour." *J. Agric. Food Chem.*54: 1296-1301 (2006).